





Short stereocontrolled synthesis of (2S,3S,4R)-3,4-dihydroxyglutamic acid

Nicole Langlois *

Institut de Chimie des Substances Naturelles, C.N.R.S., 91198 Gif-sur-Yvette Cedex, France

Received 10 September 1999; accepted 30 September 1999

Abstract

The first enantioselective synthesis of (2S,3S,4R)-3,4-dihydroxyglutamic acid was efficiently achieved in six steps from (3R,4R,5R)-1-tert-butoxycarbonyl-3,4-epoxy-5-(1-ethoxy)ethoxymethyl pyrrolidin-2-one 4 derived from (S)-pyroglutaminol. © 1999 Elsevier Science Ltd. All rights reserved.

L-Glutamic acid 1 plays a crucial role as excitatory neurotransmitter in the mammalian central nervous system. It is implicated in several neurological disorders and it has, accordingly, generated extensive interest. Many derivatives of this aminodiacid, 1-3 as for instances 4-alkylated analogues, 4 have been synthesized in order to modulate its biological activities, but 4-hydroxylated derivatives have been less studied. Few 3,4-disubstituted derivatives have been described and, while (2S)-3,4-dihydroxyglutamic acid has been isolated from *Lepidium sativum* and *Rheum rhaponticum*, 6 the 3S,4R diastereomer 2 has never been synthesized. A synthesis of the 3S,4S diastereomer 3 has been reported recently. We describe here the stereocontrolled synthesis of 2.

A stereoselective route to highly functionalized hydroxy-oxazolidin-2-ones of definite configurations was recently developed in our laboratory through intramolecular ring opening of epoxides with the mediation of neighbouring tert-butyl carbamates. These results suggested a straightforward pathway to the (2S,3S,4R)-3,4-dihydroxyglutamic acid 2, in which an oxazolidinone ring served as β-aminoalcohol protective group (Scheme 1). Thus, the opening of the epoxide 5 (derived, through 4, from (S)-pyroglutaminol) with participation of the vicinal N-Boc group occurred cleanly when 5 was adsorbed on silica gel and quantitatively afforded the oxazolidinone 6.8 The secondary alcohol of 6 was protected as tert-butyldimethylsilylether according to a classical protocol (7, 90%) and the primary alcohol function

^{*} Fax: 01-69 07 72 47; e-mail: nicole.langlois@icsn.cnrs-gif.fr

was selectively deprotected smoothly under mild acid conditions to give 8 (97%). Attempts to oxidize 8 into the corresponding carboxylic acid, with sodium periodate in excess and catalytic ruthenium trichloride (CCl₄-CH₃CN-H₂O, rt) did not lead to the expected result, although this oxidizing system has been widely used with polyfunctionalized molecules. The oxidation of 8 with Jones reagent was efficient. It kept the *tert*-butyldimethylsilyloxy group unaltered, giving rise to the carboxylic acid 9 which gave broad signals in H NMR and, for this reason, was fully characterized as its methyl ester 10 (70% for two steps, not optimized).

Scheme 1.

As natural (2S)-3,4-dihydroxyglutamic acid is known to be unaffected by strong acidic conditions, 6a all the protecting groups of the dimethylester 10 were removed in one pot by heating at 80°C in 6N HCl. 10,11 The crude aminodiacid 2 was isolated as its hydrochloride by evaporation at 40°C of the reaction mixture, followed by precipitation of the residue in H_2O -EtOH-Et₂O (88%). Its mass spectrum (FAB) exhibited a peak (M+H)⁺ at m/z 180, and NMR spectra indicated no cyclization to the dihydroxypyroglutamic acid $11.^{12}$

Thus, the first enantioselective synthesis of (2S,3S,4R)-3,4-dihydroxyglutamic acid was efficiently achieved from the epoxide 4.

References

- (a) Excitatory Amino Acids Receptors: Design of Agonists and Antagonists, Krogsgaard-Larsen, P.; Hansen J. J. Eds.; Ellis Horwood: Chichester, 1992. (b) Rojas, A. Ph.D. Thesis, 1994, University of Paris XI, Orsay, Order no. 3264, 1994. (c) Moloney, M. G. Nat. Prod. Rep. 1999, 16, 485-498.
- 2. Watkins, J. C.; Krogsgaard-Larsen, P.; Honoré, T. Trends Pharmacol. Sci. 1990, 11, 25-33.
- 3. See, for examples: (a) Shimamoto, K.; Ohfune, Y. J. Med. Chem. 1996, 39, 407-423, and references cited therein. (b) Conti, P.; Dallanoce, C.; De Amici, M.; De Micheli, C.; Fruttero, R. Tetrahedron 1999, 55, 5623-5634.
- (a) Righini-Tapie, A.; Azerad, R. J. Appl. Biochem. 1984, 6, 361-366. (b) Belokon, Y. N.; Bulychev, A. G.; Ryzhov, M. G.; Vitt, S. V.; Batsanov, A. S.; Struchkov, Y. T.; Bakhmutov, V. I. Belikov, V. M. J. Chem. Soc., Perkin Trans. 1 1986, 1865-1872. (c) Baldwin, J. E.; Adlington, R. M.; Robinson, N. G. J. Chem. Soc., Chem. Commun. 1987, 153-155. (d) Echalier, F.; Constant, O.; Bolte, J. J. Org. Chem. 1993, 58, 2747-2750. (e) Moody, C. M.; Young, D. W. Tetrahedron Lett. 1994, 35, 7277-7280. (f) Gu, Z.-Q.; Hesson, D. P.; Pelletier, J. C.; Maccecchini, M. L.; Zhou, L.-M.; Skolnick, P. J. Med. Chem. 1995, 38, 2518-2520. (g) Wermuth, C. G.; Mann, A.; Schoenfelder, A.; Wright, R. A.; Johnson, B. G.; Burnett, J. P.; Mayne, N. G.; Schoepp, D. D. J. Med. Chem. 1996, 39, 814-816. (h) Helaine, V.; Rossi, J.; Bolte, J. Tetrahedron Lett. 1999, 40, 6577-6580.
- (a) Benoiton, L.; Winitz, M.; Birnbaum, S. M.; Greenstein, J. P. J. Am. Chem. Soc. 1957, 79, 6192-6198.
 (b) Kristensen, E. P.; Larsen, M.; Olsen, O.; Sørensen, H. Acta Chem. Scand. 1980, B34, 497-504.
 (c) Passerat, N.; Bolte, J. Tetrahedron Lett. 1987, 28, 1277-1280.
 (d) Kusumi, T.; Kakisawa, H.; Suzuki, S.; Harada, K.; Kashima, C. Bull. Chem. Soc. Jpn. 1978, 51, 1261-1262.
 (e) Hanessian, S.; Vanasse, B. Can. J. Chem. 1993, 71, 1401-1406.

- (a) Virtanen, A. I.; Ettala, T. Acta Chem. Scand. 1957, 11, 182–184.
 (b) Müller, A. L.; Uusheimo, K. Acta Chem. Scand. 1965. 19. 1987–1988.
- 7. Dauban, P.; de Saint-Fuscien, C.; Dodd, R. H. Tetrahedron 1999, 55, 7589-7600.
- 8. (a) Langlois, N.; Moro, A. XVIIIth European Colloquium on Heterocyclic Chemistry, Oct. 1998, Rouen, France. (b) Langlois, N.; Moro, A. Eur. J. Org. Chem., in press.
- 9. (a) Martin, V. S.; Palazon, J. M.; Rodriguez, C. M. In *Encyclopedia of Reagents for Organic Synthesis*; Paquette, L. A., Ed.; 1995; Vol. 6, pp. 4415-4422, and references cited therein.
- 10. Delle Monache, G.; Di Giovanni, M. C.; Misiti, D.; Zappia, G. Tetrahedron: Asymmetry 1997, 8, 231-243.
- 11. It was thought that these conditions would be suitable because (2S,4R)-4-hydroxyglutamic acid is not sensitive to lactonization in acidic media, compared with the (2S,4S)-diastereomer.^{5a-d} 3,4-Dihydroxypyroglutamic acid 11, which could constitute a by-product, would be converted to the aminodicarboxylic acid 2 in 6N HCl.^{6a}
- 12. Data for 2, HCl: $[\alpha]_D^{28}$ =+8.5 (c 1.08, 18% HCl). MS (FAB) m/z=180 (M+H)⁺. ¹H NMR [300 MHz, D₂O, HOD: δ =4.80 ppm, J (hertz)]: 4.36 (d, 1H, J=3.6), 4.65 (d, 1H, J=1.5), 4.69 (m, 1H); ¹³C NMR (75.0 MHz, D₂O, dioxane: δ =67.34 ppm): 175.3 (CO), 170.9 (CO), 73.5 (CH), 69.1 (CH), 56.9 (CH).